

- New York, N.Y., 1962; H. H. Ussing, P. Kruhoffer, J. H. Thaysen, and N. A. Thorn, *The Alkali Metal Ions in Biology*, *Handb. Exp. Pharmacol.*, **13** (1960).
- (3) A. S. Perlin, P. H. du Penhoat, and H. S. Isbell, *Adv. Chem. Ser.*, **No. 117**, 39 (1973).
- (4) More than 85% according to the  $^{13}\text{C}$  spectrum assuming equal  $T_1\rho$  values for equivalent carbons in the two anomeric forms. This anomeric distribution is not significantly changed upon addition of sodium perchlorate at the concentrations in this study.
- (5) S. J. Angyal, *Tetrahedron*, **30**, 1695 (1974).
- (6) J. Andrasko and S. Forsen, *Biochem. Biophys. Res. Commun.*, **52**, 233 (1973).
- (7) C. Detellier and P. Laszlo, to be submitted for publication; see also A. T. Tsatsas, R. W. Stearns, and W. M. Risey, *J. Am. Chem. Soc.*, **94**, 5247 (1972).
- (8) It was not possible to perform an independent check from the sodium-23 chemical shift; within experimental error, it remains invariant in the presence of sugars at the concentrations used in this study.
- (9) Using a trigonal bipyramidal idealized geometry, with equatorial oxygens at 2.55 Å, and nitrogens at 2.65 Å, CNDO-2 charges of 0.25 (O) and 0.14 (N) lead to a reasonable<sup>10</sup> value of ca. 1 MHz for the quadrupolar coupling constant. This value could be somewhat higher if one were dealing with a sugar-bridged  $\text{Na}^+\text{ClO}_4^-$  ion pair, bringing a perchlorate oxygen into van der Waals contact of the cation.
- (10) D. H. Haynes, B. C. Pressman, and A. Kowalsky, *Biochemistry*, **10**, 852 (1971).
- (11) J. P. Kintzinger and J. M. Lehn, *Mol. Phys.*, **22**, 273 (1971).
- (12) P. G. Gertenbach and A. I. Popov, *J. Am. Chem. Soc.*, **97**, 4738 (1975).
- (13) Note Added in Proof. The sodium-23 line broadenings obtained with monohydric alcohols and diols such as cyclohexane-1,1-dimethanol are negligible by comparison to those observed with sugars. Indeed the interaction is specific of sugars.

Christian Detellier, Jean Grandjean, Pierre Laszlo\*

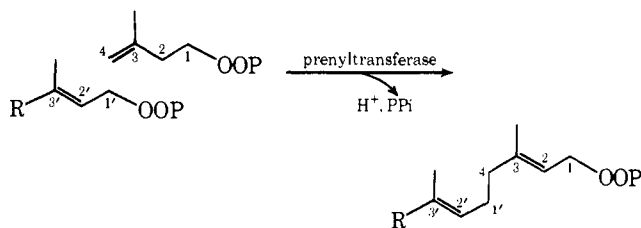
*Institut de Chimie, Université de Liège  
Sart-Tilman par 4000 Liège, Belgium*

Received January 27, 1976

## Prenyltransferase. The Mechanism of the Reaction<sup>1</sup>

Sir:

Prenyltransferase (EC 2.5.1.1) catalyzes the condensation between  $\text{C}_4$  of isopentenyl pyrophosphate (IPP) and  $\text{C}_1'$  of an allylic pyrophosphate, giving the five-carbon homologue of the allylic pyrophosphate. This is the fundamental chain elongation

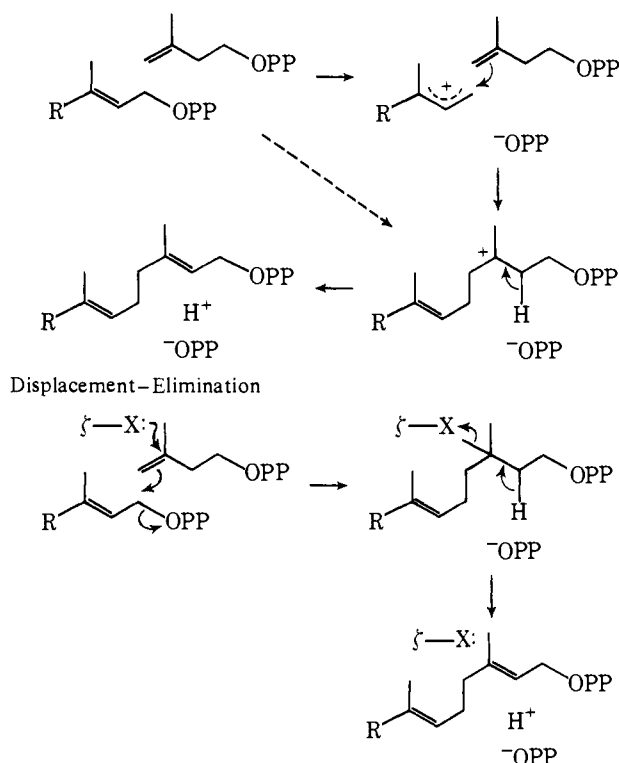


**Table I.** Effect of Methyl and Trifluoromethyl Groups

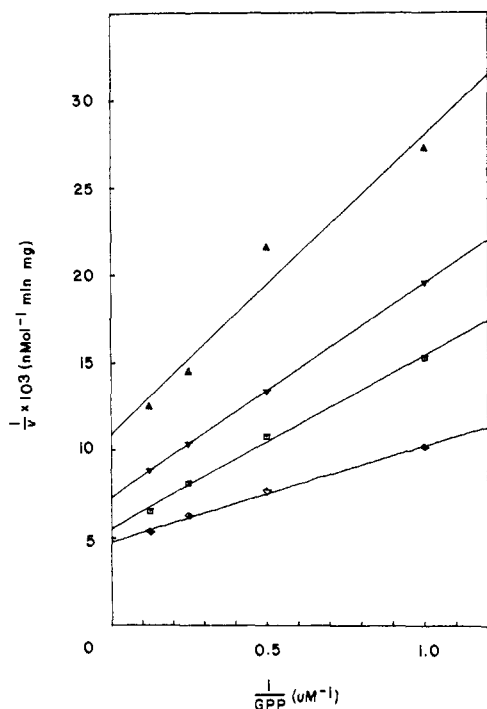
Reactant	$T$ , °C	Solvent (% acetone- $\text{H}_2\text{O}$ )	$k$	$k_{\text{CF}_3/\text{CH}_3}$
SN1 <i>E</i> -3-Trifluoromethyl-2-buten-1-yl methanesulfonate ( <i>E</i> -1-OMs)	20	92	$5.5 \times 10^{-8}$ <i>a-c</i>	$1.8 \times 10^{-6}$
	20	50	$1.55 \times 10^{-6}$ <i>a,c</i>	
	60	50	$1.28 \pm 0.03 \times 10^{-4}$ <i>c</i>	
	70	50	$3.06 \pm 0.20 \times 10^{-4}$ <i>c</i>	
	80	50	$7.98 \pm 0.55 \times 10^{-4}$ <i>c</i>	
3-Methyl-2-buten-1-yl methanesulfonate (4-OMs) <sup>d</sup>	0	92	$3.82 \pm 0.11 \times 10^{-3}$ <i>c</i>	
	20	92	$2.99 \pm 0.12 \times 10^{-2}$ <i>c</i>	
SN2 <i>E</i> -1-Chloro-4,4,4-trifluoro-2-butene (5)	20	100	$68.8 \times 10^{-5}$ <i>e,f</i>	11
	20	100	$6.1 \times 10^{-5}$ <i>e,g</i>	

<sup>a</sup> Extrapolated from higher temperatures.  $\Delta H^\ddagger = 20.7 \text{ kcal mol}^{-1}$ ,  $\Delta S^\ddagger = -15 \text{ eu}$ . <sup>b</sup> Extrapolated 50% acetone-water,  $m = 0.383$ ,  $Y = 2.25$ . <sup>c</sup> Units are  $\text{s}^{-1}$ . <sup>d</sup>  $\Delta H^\ddagger = 15.8 \text{ kcal mol}^{-1}$ ,  $\Delta S^\ddagger = -12 \text{ eu}$ . <sup>e</sup> For displacement with  $\text{I}^-(\text{KI})$ ,  $1. \text{ mol}^{-1} \text{ s}^{-1}$ . <sup>f</sup> E. T. McBee, R. D. Battershell, and H. P. Braendlin, *J. Am. Chem. Soc.*, **84**, 3157 (1962); J. A. Pegolotti and W. G. Young, *ibid.*, **83**, 3258 (1961). <sup>g</sup> L. F. Hatch and S. S. Nesbitt, *ibid.*, **73**, 358 (1951).

**Scheme I.** Ionization-Condensation-Elimination



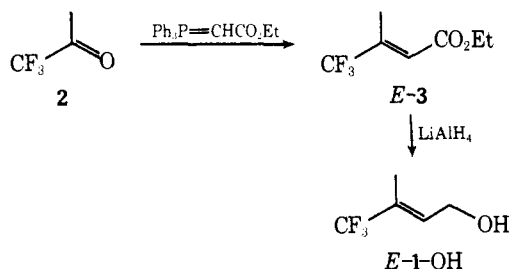
reaction of terpene biosynthesis and leads to the formation of several important classes of natural products such as sterols, carotenoids, dolichols, and respiratory coenzymes. The mechanisms which have been proposed for prenyl transfer can be grouped into two broad categories (Scheme I): those in which condensation is initiated by heterolytic cleavage of the carbon-oxygen bond, with or without assistance from the double bond of isopentenyl pyrophosphate, yielding cationic intermediates (ionization-condensation-elimination),<sup>2</sup> and those in which condensation is initiated by attack of a nucleophilic group at the double bond of isopentenyl pyrophosphate with simultaneous formation of the  $\text{C}_1'$ - $\text{C}_4$  bond between the two substrates and rupture of the  $\text{C}_1'$ -oxygen bond (displacement-elimination).<sup>2d,3</sup> We reasoned that it would be possible to distinguish between the two mechanisms by replacing the R group in the allylic substrate by a trifluoromethyl group. The strong electron-withdrawing effect of a trifluoromethyl substituent ( $\sigma^+ = 0.612$ )<sup>4</sup> should retard the rate of



**Figure 1.** Inhibition of prenyltransferase by *E*-1-OPP. Incubations were at 37° in 200  $\mu$ l of 10 mM potassium phosphate buffer, pH 7.0, 1 mM  $\text{MgCl}_2$ , 0.1 mM dithiothreitol, and 2  $\mu$ M isopentenyl pyrophosphate with 69 ng of prenyltransferase (specific activity 250) and the indicated concentrations of geranyl pyrophosphate. Determinations were in duplicate. Concentration of *E*-1-OPP;  $\diamond$  none,  $\square$  50  $\mu$ M,  $\nabla$  100  $\mu$ M,  $\Delta$  200  $\mu$ M.

ionization of the allylic pyrophosphate while having little effect on the rate of a direct nucleophilic displacement at  $\text{C}_1'$ . In this communication, we present preliminary results with an allylic substrate analogue, *E*-trifluoromethyl-2-butenyl pyrophosphate (*E*-1-OPP).

*E*-3-Trifluoromethyl-2-buten-1-ol (*E*-1-OH) was prepared by the sequence of reactions shown below.



The Wittig condensation<sup>5</sup> gave a 97:3 mixture of *E*<sup>6</sup> and *Z*<sup>7</sup> isomers which were separated by spinning band distillation. The stereochemistry of the double bond was established by a nuclear Overhauser experiment in which irradiation of the protons at  $\text{C}_4$  produced a  $32 \pm 3\%$  enhancement of the olefinic proton in *Z*-3 and no enhancement,  $3 \pm 3\%$ , for the *E* isomer. Alcohol *E*-1-OH,<sup>8</sup> obtained by hydride reduction at 0°, was converted into methanesulfonate (*E*-1-OMs) and pyrophosphate<sup>9,10</sup> (*E*-1-OPP) derivatives for chemical and enzymatic studies.

The data in Table I illustrate the effect of replacing a methyl group by trifluoromethyl in typical  $\text{S}_\text{N}1$  (solvolysis in aqueous acetone) and  $\text{S}_\text{N}2$  (displacement by iodide in dry acetone) reactions for allylic systems. It is clear that the trifluoromethyl moiety severely retards heterolytic cleavage of the  $\text{C}_1$ -oxygen bond in *E*-1-OMs and mildly accelerates direct nucleophilic substitution at an allylic center.

When *E*-1-OPP is incubated<sup>11</sup> with isopentenyl pyrophos-

phate and prenyltransferase from porcine liver,<sup>12</sup> the rate of condensation is  $5.1 \times 10^{-4}$  nmol/(min mg).<sup>13</sup> In comparison, the rate observed with isopentenyl pyrophosphate and geranyl pyrophosphate is  $7.4 \times 10^2$  nmol/(min mg)<sup>14</sup> under similar conditions. Assuming that the rate of the reaction varies linearly with the concentration of enzyme, *E*-1-OPP is at least  $1.5 \times 10^6$  times less reactive than geranyl pyrophosphate.

Inefficient utilization of the trifluoromethyl analogue cannot be attributed to poor binding with prenyltransferase. The size of the trifluoromethyl group is similar to that of a methyl group,<sup>15</sup> and the enzyme catalyzes condensation between isopentenyl pyrophosphate and a variety of allylic pyrophosphates with different alkyl substituents at  $\text{C}_3'$ .<sup>16</sup> Also, kinetic studies summarized in Figure 1 show that *E*-1-OPP binds to the enzyme.<sup>17</sup>

We conclude that this condensation takes place by an ionization-condensation-elimination mechanism. The exact timing of condensation with regard to ionization cannot be firmly established although two observations suggest the possibility that condensation lags behind ionization. The trifluoromethyl moiety retards the rates for solvolysis and prenyl transfer to a similar extent, implying substantial delocalization of positive charge into the  $\text{C}_2'$ - $\text{C}_3'$  double bond during ionization for both reactions. Also, we have recently shown that prenyltransferase catalyzes hydrolysis of its allylic substrate in the absence of isopentenyl pyrophosphate.<sup>18</sup> Experiments are underway in our laboratories which will further delineate the mechanism of prenyltransferase.

## References and Notes

- (1) This investigation was supported by research grants GM 21328 and AM 13140 from the National Institutes of Health.
- (2) (a) F. Lynen, H. Eggerer, U. Henning, and I. Kessel, *Angew. Chem.*, **70**, 738 (1958); (b) H. C. Rilling and K. Bloch, *J. Biol. Chem.*, **234**, 1424 (1959); (c) J. W. Cornforth and G. Popjak, *Tetrahedron Lett.*, No. 19, 29 (1959); (d) J. W. Cornforth, *Angew. Chem., Int. Ed. Engl.*, **7**, 903 (1968).
- (3) J. W. Cornforth, R. H. Cornforth, G. Popjak, and L. Yengoyan, *J. Biol. Chem.*, **241**, 3970 (1966).
- (4) Y. Okamoto, T. Inukai, and H. C. Brown, *J. Am. Chem. Soc.*, **80**, 4969 (1958).
- (5) D. L. Dull, I. Baxter, and H. S. Mosher, *J. Org. Chem.*, **32**, 1622 (1967).
- (6) NMR ( $\text{CDCl}_3$ ,  $\text{Me}_4\text{Si}$  internal standard)  $\delta$  1.30 (3, t,  $J = 7$  Hz,  $\text{CH}_3$  of ethyl group), 2.24 (3, d,  $J = 1.6$  Hz, H at  $\text{C}_4$ ), 4.21 (2, q,  $\text{CH}_2$  of ethyl group), and 6.21 ppm (1, septet, H at  $\text{C}_2$ ).
- (7) NMR ( $\text{CDCl}_3$ ,  $\text{Me}_4\text{Si}$  internal standard)  $\delta$  1.32 (3, t,  $J = 7$  Hz,  $\text{CH}_3$  of ethyl group), 2.00 (3, d,  $J = 1.6$  Hz, H at  $\text{C}_4$ ), 4.21 (2, q,  $\text{CH}_2$  of ethyl group), and 6.06 ppm (1, q, H at  $\text{C}_2$ ).
- (8) E. T. McBee, O. R. Pierce, and D. D. Smith, *J. Am. Chem. Soc.*, **76**, 3725 (1954).
- (9) P. W. Holloway and G. Popjak, *Biochem. J.*, **104**, 57 (1967).
- (10) *E*-1-OPP gave a single spot on TLC and contained 2 equiv of phosphate. O. C. Richards and P. D. Boyer, *J. Mol. Biol.*, **11**, 327 (1965).
- (11) All incubations were at 37° in a mixture of 10 mM potassium phosphate buffer, pH 7.0, and 0.1 mM dithiothreitol. Total volume was 200  $\mu$ l. The acid-lability method was used for all assays. B. C. Reed and H. C. Rilling, *Biochemistry*, **14**, 50 (1975).
- (12) The procedure for isolation of prenyltransferase from porcine liver will be published elsewhere.
- (13) 410 nmol of *E*-1-OPP, 8 nmol of IPP, and 35  $\mu$ g of enzyme, specific activity 740. After 4033 min, the activity of the enzyme had only dropped by 10%.
- (14) 8 nmol of GPP, 8 nmol of IPP, and 35 ng of enzyme, specific activity 740
- (15) (a) W. F. Edgell, G. B. Miller, and J. W. Amy, *J. Am. Chem. Soc.*, **79**, 2391 (1957); (b) J. L. DeCoen, G. Elefante, A. M. Liquori, and A. Damiani, *Nature (London)*, **216**, 910 (1967).
- (16) (a) T. Nishino, K. Ogura, and S. Seto, *Biochim. Biophys. Acta*, **302**, 33 (1973); (b) T. Nishino, K. Ogura, and S. Seto, *J. Am. Chem. Soc.*, **94**, 6849 (1972); (c) T. Nishino, K. Ogura, and S. Seto, *ibid.*, **93**, 794 (1971); (d) K. Ogura, T. Nishino, T. Koyama, and S. Seto, *ibid.*, **92**, 6036 (1970); (e) G. Popjak, J. L. Rabinowitz, and J. M. Baron, *Biochem. J.*, **113**, 861 (1969).
- (17) Equilibrium dialysis experiments with crystalline prenyltransferase indicate that the two catalytic sites in the dimer (one per subunit) are identical. B. C. Reed, Ph.D. Dissertation, University of Utah, 1976.
- (18) C. D. Poulter and H. C. Rilling, *Biochemistry*, **15**, 1079 (1976).
- (19) Alfred P. Sloan Fellow.
- (20) Career Development Award from the National Institutes of Health, HL 00084.

C. Dale Poulter,<sup>\*19,20</sup> Dennis M. Satterwhite, Hans C. Rilling\*  
 Departments of Chemistry and Biochemistry  
 University of Utah  
 Salt Lake City, Utah 84112  
 Received December 22, 1975